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(54) Title: REPLICATION INCOMPETENT HERPES VIRUS VECTORS

(57) Abstract: Use replication incompetent herpes virus comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system disorder by administering said medicament to a peripheral nerve, in a method of determining whether a gene has an effect on a phenotype associated with a peripheral nervous system disorder and in a method of treatment of a disorder of the peripheral nervous system.

WO 01/46450 A1

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REPLICATION INCOMPETENT HERPES VIRUS VECTORS

Field of the Invention

The present invention relates to replication incompetent herpes simplex
5 viruses capable of efficiently transferring genes to cells of the peripheral nervous
system. It also relates to the use of such viruses in the study and treatment of diseases
and conditions of the nervous system.

Background of the Invention

10 Herpes simplex viruses 1 and 2 are large DNA viruses the lifecycles of which
are characterised by the ability to enter a lifelong latent state in the sensory ganglia of
an infected individual from which reactivation of the virus can occur intermittently.
During latency the HSV genome, which remains extra-chromosomal, is largely
quiescent although the fact that a small part of the genome encoding the latency
15 associated transcripts (LATs) is transcribed during latency (Stevens, 1987) shows
that the development of gene delivery vectors allowing transgene expression during
this time should be possible. During latency HSV is not naturally cleared by the
immune system and infected cells remain undamaged, further showing the potential
for a long-term therapeutic benefit using HSV as a vector (reviewed by Coffin and
20 Latchman 1995, Fink 1996). HSV also has the unique ability among viruses currently
under development as vectors to infect axonal nerve terminals followed by efficient
retrograde axonal transport of the virus to the cell body in the spinal ganglia. Thus
HSV vectors might be developed allowing peripheral vector inoculation to give gene
delivery to the spinal ganglia following such retrograde transport. There are a number
25 of potential applications for vectors capable of axonal transport in the peripheral
nervous system, including the stimulation of regrowth of damaged nerves, the
study/treatment of various pain states, the protection of neurons from further
degeneration in e.g. motor neuron disease, the study/treatment of various
neuropathies, the study of neuronal development, and the screening of the relevance
30 of genes implicated as being important in any of these processes by a gene delivery
approach.

Previous work has shown that HSV vectors can deliver genes to spinal

ganglia by such routes, but this in each case has used vectors retaining at least same replication competence as this has been thought necessary for effective gene delivery. Here various mutant viruses have been used variously mutated for the non-essential genes encoding ICP6 (Goldstein and Weller 1988), ICP0 (Leib *et al.* 1989), TK
5 (Palella *et al.* 1988, Ho and Mokarski 1988), gC (Lokensgard *et al.* 1994; Dobson *et al.* 1995; Goins *et al.* 1994), vmw65 (Ecob Prince *et al.* 1995), un-disabled (Margolis *et al.* 1993, Lachman *et al.* 1996, Lachmann and Efstathiou, 1997), or deleted for ICP34.5 or ICP34.5 and vmw65 (Coffin *et al.* 1996). Each of these viruses is attenuated compared to wild type virus even though the mutated genes are non-
10 essential, and each have shown reasonable levels of at least short term gene delivery and/or the ability to enter latency following inoculation of animals by the footpad or ear pinna routes. As discussed however, all retain at least same degree of replication competence in vivo and in culture as no essential genes have been inactivated. Therefore, while replication incompetent HSV vectors have been available for some
15 time, none have been used in the peripheral nervous system as it has so far been assumed that viruses incapable of replication in any cell (i.e. mutated for an essential gene/genes) would not allow gene delivery to spinal ganglia following inoculation by peripheral routes.

20 Summary of the Invention

We have previously reported that HSV vectors mutated for ICP34.5 or ICP34.5 and vmw65 can give gene delivery to the peripheral nervous system following footpad inoculation (Coffin *et al.*, 1996). However, similar to other work discussed above, such viruses while attenuated still retain some degree of replication
25 competence *in vivo*. Indeed it had been assumed that such replication competence might be necessary for gene delivery by such a route as the virus is required to traffic from the point of inoculation to the cell body in the spine where the delivered gene is to be expressed. Replication might be imagined to be necessary for this to occur efficiently. Therefore while promising results of gene delivery to the peripheral
30 nervous system have been reported, including the alteration of sensitivity to painful stimuli by an HSV-vector mediated approach (Wilson *et al.* 1999), the vectors used retain replication competence raising issues of safety and acceptability if they are to

be used for human treatment. When such viruses carry an exogenous gene or genes, issues of safety and acceptability may also arise when they are used in laboratory based pre-clinical and basic research. The present invention describes vectors which are highly efficient at gene delivery to the peripheral nervous system following peripheral inoculation and which have a considerably improved safety/acceptability profile over vectors which have previously been available for such a purpose.

In summary, we have made the surprising finding that highly disabled HSV vectors which cannot replicate in any cell other than those used to prepare vector stocks (i.e. essential gene/genes have been inactivated in the vector which are complemented in the producer cell line) can give highly efficient gene delivery to the peripheral nervous system following direct injection into a peripheral nerve. Such a result, giving very high efficiency gene delivery, has not previously been reported and thus HSV vectors which show an improved safety/acceptability profile over those which have previously been used in the peripheral nervous system are provided by the current invention. Such replication incompetent herpes virus vectors may be used in methods of studying or treating disorders of the peripheral nervous system. In particular, such vectors may be used in methods of target validation, for examples by screening genes to identify what, if any, role they play in a peripheral nervous system disorder.

Accordingly, the present invention provides:

- use of a replication incompetent herpes virus comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system disorder by administering said medicament to a peripheral nerve;
- method of treating a subject suffering from a disorder of the peripheral nervous system or of preventing a peripheral nervous system disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of a replication incompetent herpes virus into a peripheral nerve of the subject;
- method of determining whether a gene has an effect on a phenotype associated with a peripheral nervous system disorder or on a cell of the peripheral nervous system which is relevant to a peripheral nervous system disorder, which method comprises:

- (i) inoculating a replication incompetent herpes virus comprising a

heterologous gene into a peripheral nerve; and

(ii) monitoring a phenotype of said disorder or an effect of expression of said gene on said cell to determine thereby whether said gene has an effect relevant to said disorder;

- 5 - a method of screening genes implicated in a peripheral nervous system disorder to identify a target for gene therapy or small molecule modulators, which method comprises a method according to the invention.

Detailed Description of the Invention

10 Viruses

The present invention provides a method for identifying a gene that may be used as a target for therapeutic intervention in the treatment or prevention of a peripheral nervous system disorder. Such genes are identified by expressing a test gene in a cell of the peripheral nervous system using a replication incompetent herpes virus vector to deliver the gene to the cell and by determining whether expression of the gene affects a phenotype associated with a peripheral nerve system disorder. A method of determining whether a gene influences a phenotype associated with a peripheral nerve system disorder consists essentially of the following steps:

- (i) inoculating a peripheral nerve *in vivo* or *in vitro* with a replication
20 incompetent herpes virus capable of efficiently infecting a cell of the peripheral nervous system, which virus comprises a test gene;
- (ii) monitoring a phenotype of said disorder to determine thereby whether expression of said gene has an effect on said phenotype.

The present invention provides a method for screening genes to identify a
25 gene which is important in a peripheral nervous system disorder and which may be used as a target for identifying agents for treating the disorder.

The present invention also provides the use of a replication incompetent herpes virus vector in the manufacture of a medicament for the treatment of a peripheral nervous system disorder. A method of treating a subject suffering from
30 peripheral nervous system disorder by administering a therapeutically effective amount of a replication incompetent herpes virus vector to the peripheral nervous system of a subject in need thereof is also provided by the present invention. The

herpes virus vector is typically replication incompetent and is capable of efficiently infecting a cell of the peripheral nervous system. Preferably the herpes virus vector comprises a heterologous gene. The heterologous gene typically comprises a polypeptide of therapeutic use.

- 5 Administration of the herpes virus vector to a subject suffering from disorder of the peripheral nervous system typically alleviates the symptoms of the disorder and/or prevents the progression of the disorder.

A virus for use in the invention is capable of efficiently infecting a cell of the peripheral nervous system following peripheral inoculation, but is incapable of
10 replicating in any cell unless at least one gene which has been rendered non-functional in the virus is also expressed in the cell, i.e. the virus is replication incompetent other than in the cells in which virus stocks are prepared. A virus is replication incompetent if it is incapable of replicating in a cell which does not express at least one functional essential viral gene.

- 15 A virus capable of efficiently infecting a cell of the peripheral nervous system following peripheral inoculation typically infects at least 50 cells for example, at least 60, at least 70, at least 80, at least 90 or at least 100 cells, more preferably at least 200, for example at least 500 or at least 1000 cells following direct injection of virus at a titre of from $1 \times 1 \times 10^8$ pfu/ml to 1×10^9 pfu/ml, preferably from 3×10^8
20 pfu/ml to 7×10^8 pfu/ml, more preferably from 4×10^8 pfu/ml to 6×10^8 pfu/ml, more preferably 5×10^8 pfu/ml into a peripheral nerve, for example the sciatic nerve, of a laboratory animal. This can be determined by monitoring the expression heterologous marker gene such as LacZ or GFP operably linked to a control sequence, preferably a control sequence which is active during herpes virus latency.
25 Preferably the peripheral nervous system cells are dorsal root ganglion (DRG) cells. Preferably the laboratory animal is a primate or a rodent such as a rat or a mouse.

- By definition such viruses are attenuated such that they are replication incompetent. A virus may be rendered replication incompetent by the mutation of at least one essential gene such that the essential gene is non-functional. Viral regions
30 altered for the purposes of such attenuation may be eliminated (completely or partly), made non-functional, or substituted by other sequences, in particular by a heterologous gene sequence. Attenuating mutations resulting in a replication

incompetent virus have been described for all neurotrophic herpes viruses so far described. Replication incompetent viruses are generally capable of replicating in cell lines which express a functional essential viral gene in which compensates for the absence of a functional essential viral gene in the replication incompetent virus.

- 5 Such cell lines may be used to prepare viral stocks.

Although the present invention has been exemplified using herpes simplex viruses, it will be understood that other neurotrophic viruses of the herpesviridae family may be modified such that they are replication incompetent and can achieve efficient gene delivery to cells of the peripheral nervous system following peripheral
10 inoculation. In particular, such viruses may include varicella zoster virus, pseudo-rabies virus or bovine herpes viruses. In addition, herpes amplicon vectors are replication incompetent herpesviruses. Thus, herpes amplicon vector stocks which are capable of infecting cells of the peripheral nervous system following peripheral nerve inoculation may also be used in accordance with the present invention.

- 15 When the virus of the invention is a herpes simplex virus, the virus may be derived from, for example, HSV1 or HSV2 strains, or derivatives thereof, preferably HSV1. Derivatives include inter-type recombinants containing DNA from HSV1 and HSV2 strains. Such inter-type recombinants are described in the art, for example in Thompson *et al.* 1998 and Meignier *et al.* 1988. Derivatives preferably have at least
20 70% sequence homology to either the HSV1 or HSV2 genomes, more preferably at least 80%, even more preferably at least 90 or 95% and most preferably at least 96, 97, 98 or 99%. More preferably derivatives have at least 70% sequence identity to either a HSV1 or HSV2 genome, more preferably at least 80%, even more preferably at least 90 or 95% and most preferably at least 96, 97, 98 or 99%.

- 25 A derivative may have the sequence of a HSV1 or HSV2 genome modified by nucleotide substitutions, for example from 1, 2 or 3 to 10, 25, 50 or 100 substitutions. The HSV1 or HSV2 genome may alternatively or additionally be modified by one or more insertions and/or deletions and/or by an extension at either or both ends.

- 30 For example the UWGCG Package provides the BESTFIT program which can be used to calculate homology (for example used on its default settings) (Devereux *et al.* (1984) *Nucleic Acids Research* 12, p387-395). The PILEUP and

BLAST algorithms can be used to calculate homology or line up sequences (typically on their default settings), for example as described in Altschul (1993) *J. Mol. Evol.* 36:290-300; Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-10.

Software for performing BLAST analyses is publicly available through the
5 National Centre for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pair (HSPs) by identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold
10 (Altschul *et al.*, 1990). These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Extensions for the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved
15 value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLAST program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1992) *Proc. Natl. Acad.*
20 *Sci. USA* 89: 10915-10919) alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of both strands.

The BLAST algorithm performs a statistical analysis of the similarity between two sequences; see e.g., Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90: 5873-5787. One measure of similarity provided by the BLAST algorithm
25 is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a sequence is considered similar to another sequence if the smallest sum probability in comparison of the first sequence to the second sequence is less than about 1, preferably less than about 0.1, more preferably less
30 than about 0.01, and most preferably less than about 0.001.

Viruses of the invention may be functionally inactive for individual (e.g. ICP27 or ICP4) or multiple (ICP27 and/or ICP4 and/or ICP22 and/or ICP0 and/or

ICP47) immediate early genes. They may additionally contain a mutation in the gene encoding *vmw65* reducing/preventing its transcriptional activation capability. Alternatively essential genes which are not immediate early genes, such as essential genes involved in virus replication, for example the products of UL29 and UL30 genes, single stranded DNA binding protein and DNA polymerase respectively, may be deleted either alone, in any combination, or in combination also with the inactivation of one or more immediate early genes and/or *vmw65*. Examples of such viruses include viruses reported by Samaniego *et al.* 1998, Krisky *et al.* 1998, or Thomas *et al.* 1999. Particularly preferred viruses are those in which immediate early gene expression has been prevented or minimised for example, a virus with inactivating mutations in or deletions of the genes encoding ICP4, ICP27, ICP0 and ICP22 is preferred. Viruses with inactivating mutations in or deletions of ICP27 and/or ICP4 and also such a virus with a mutation in *vmw65* reducing/preventing its trans-activating activity are also preferred.

Preferably viruses of the invention do not express, or express only very low levels of at least two immediate early genes. Very low levels are levels below the detection limit of a western blot using an antibody to a protein product of an immediate early gene. A virus may comprise any suitable mutation which minimises immediate early gene expression. Preferably a virus does not express, or expresses only very low levels of at least three immediate early genes. More preferably a virus does not express, or expresses only very low levels of at least four immediate early genes. The immediate early genes which must not be expressed or be expressed at only minimal levels are ICP0, ICP4, ICP22 and ICP27. The two immediate early genes may be ICP27 and ICP4, ICP27 and ICP0, ICP27 and ICP22, ICP4 and ICP0, ICP4 and ICP22 or ICP0 and ICP22.

Combinations of gene deletions useful in accordance with the present invention are those in which mutations not only prevent replication but which also minimise or reduce toxicity of the virus. Thus a virus capable of infecting a cell of the peripheral nervous system following peripheral inoculation and with mutations in ICP4 and /or ICP27 is particularly preferred. These mutations prevent expression of not only a functional form of the mutated gene itself but also other viral genes that may be cytotoxic. In addition, viruses with mutations in combinations of or in

individual essential genes which can be efficiently complemented by a cell line prepared for their growth are preferred. In particular high titres of the virus can readily be prepared. Such virus cells lines are described in the art, e.g. Samaniego et al 1998 and Krisky et al 1998. The terminology used in describing the various HSV genes is as found in Coffin and Latchman, 1996.

Production of Replication Incompetent Viruses

When the herpes simplex viruses of the invention lack a particular functional essential gene, for example a gene encoding ICP4 or ICP27, the virus of the invention is propagated on a cell line expressing that essential gene. For example, when the virus lacks a functional ICP27 gene, the virus may be propagated on V27 cells (Rice and Knipe, 1990), 2-2 cells (Smith *et al.*, 1992) or B130/2 cells (Howard *et al.* 1998). When the virus lacks a functional ICP4 gene the virus may be propagated on a cell line expressing ICP4, for example E5 cells (DeLuca *et al.*, 1985). When the virus lacks a functional ICP4 gene and a functional ICP27 gene the virus is propagated on a cell line expressing both ICP4 and ICP27 (such as E26 cells; Samaniego *et al.*, 1995), and when the virus additionally lacks a functional vmw65 gene the virus may be propagated on a cell line also containing a non-HSV homologue of vmw65 (e.g. from equine herpes virus as in Thomas *et al.* 1999). Mutations to vmw65 may also be partially compensated for by inclusion of hexamethylene bisacetamide (HMBA) in the media used for virus growth (MacFarlane *et al.* 1992).

ICP27-expressing cell lines can be produced by co-transfecting mammalian cells, for example the Vero or BHK cells, with a vector, preferably a plasmid vector, comprising a functional HSV ICP27 gene capable of being expressed in said cells, and a vector, preferably a plasmid vector, encoding a selectable marker, for example neomycin resistance. Clones possessing the selectable marker are then screened further to determine which clones also express functional ICP27, for example on the basis of their ability to support the growth of ICP27- HSV strains, using methods known to those skilled in the art (for example as described in Rice and Knipe, 1990). Cell lines which do not allow reversion of an ICP27- mutant HSV strain to a strain with functional ICP27 are produced as described above, ensuring that the vector

comprising a functional ICP27 gene does not contain sequences that overlap with (i.e. are homologous to) sequences remaining in the ICP27- mutant virus.

Where HSV strains of the invention comprise inactivating modifications in other essential genes, for example ICP4, complementing cell lines will comprise a functional HSV gene which complements the modified essential gene in the same manner as described for ICP27. For example in the case of HSV strains comprising mutations in both ICP27 and ICP4, a cell line expressing both ICP27 and ICP4 is used such as E26 cells (Samaniego *et al.*, 1995). HSV strains expressing other essential genes can be constructed in a similar manner to that described for ICP27. Here again, if it is ensured that there is no sequence overlap between the remaining virus DNA and that inserted into the cell line for virus growth, the possibility of reversion of the virus to a less disabled form during growth will be minimised.

Methods of mutation

The various viral genes referred to may be rendered functionally inactive by any suitable technique. For example, they may be rendered functionally inactive by deletions, substitutions or insertions, preferably by deletion. Deletions may remove portions of the genes or the entire gene. For example, deletion of only one nucleotide may be made, resulting in a frame shift. However, preferably larger deletions are made, for example at least 25%, more preferably at least 50% of the total coding and non-coding sequence (or alternatively, in absolute terms, at least 10 nucleotides, more preferably at least 100 nucleotides, most preferably, at least 1000 nucleotides). It is particularly preferred to remove the entire gene and some of the flanking sequences. Inserted sequences may include the heterologous genes described below. In particular, it is preferred to insert the heterologous gene into ICP27 or ICP4. In the case of the *vmw65* gene, the entire gene is not deleted since it encodes an essential structural protein, but a small inactivating mutation is made which abolishes the ability of VMW65 to activate transcriptionally IE genes (e.g. as in Ace *et al.*, 1989 or Smiley *et al.* 1997).

Mutations are made in the herpes viruses by homologous recombination methods are well known to those skilled in the art. For example, HSV genomic DNA is transfected together with a vector, preferably a plasmid vector, comprising the

mutated sequence flanked by homologous HSV sequences. The mutated sequence may comprise deletions, insertions or substitutions, all of which may be constructed by routine techniques. Insertions may include selectable marker genes, for example lacZ or GFP, for screening recombinant viruses by, for example, β -galactosidase activity or fluorescence.

Heterologous genes and promoters

The viruses for use in the invention are typically modified to carry a heterologous gene. The term "heterologous gene" encompasses any gene. A heterologous gene may be a test gene for use in a method of target validation or a gene of therapeutic use for use in a method of treatment or in the manufacture of a medicament. Although a heterologous gene is typically a gene not present in the genome of a herpes virus, a herpes gene may be used provided that the coding sequence is not operably linked to the viral control sequences with which it is naturally associated.

The heterologous gene may be any allelic variant of a wild-type gene, or it may be a mutant gene. The term "gene" is intended to cover nucleic acid sequences which are capable of being at least transcribed. Thus, sequences encoding mRNA, tRNA and rRNA are included within this definition. However, the present invention is primarily concerned with the expression of polypeptides and/or mRNA and antisense RNA rather than tRNA and rRNA. The term "gene" covers a polynucleotide comprising the coding sequence of a polypeptide. The polynucleotide may include one or more introns, for example may comprise genomic DNA. Preferably the polynucleotide comprises cDNA. Sequences encoding mRNA will optionally include some or all of 5' and/or 3' transcribed but untranslated flanking sequences naturally, or otherwise, associated with the translated coding sequence. It may optionally further include the associated transcriptional control sequences normally associated with the transcribed sequences, for example transcriptional stop signals, polyadenylation sites and downstream enhancer elements.

A heterologous gene may be inserted into the viral genome by homologous recombination of HSV strains with, for example, plasmid vectors carrying the

heterologous gene flanked by HSV sequences. The heterologous gene may be introduced into a suitable plasmid vector comprising herpes viral sequences using cloning techniques well-known in the art. The heterologous gene may be inserted into the viral genome at any location provided that the virus can still be propagated.

5 It is preferred that the heterologous gene is inserted into an essential gene. Heterologous genes may be inserted at multiple sites within the virus genome.

The transcribed sequence of a heterologous gene is preferably operably linked to a control sequence permitting expression of the heterologous gene in a mammalian cell, preferably a cell of the peripheral nervous system. The term "operably linked"

10 refers to a juxtaposition wherein the components described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is joined in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequence.

Typically the heterologous gene is under the control of a promoter active

15 during herpes virus latency. Such promoters typically comprise fragments of the latency associated transcript (LAT) region of the virus or are chimaeric promoters comprising fragments of the LAT region together with a non-LAT promoter. Such latently active promoters may drive the expression of endogenous gene or genes from within the LAT region of the virus, or when inserted at an ectopic, non-LAT site

20 within the HSV genome.

The control sequence comprises a promoter allowing expression of the heterologous gene and a signal for termination of transcription. The promoter is selected from promoters which are functional in mammalian, preferably rat, mouse, guinea pig, ferret or human, cells of the peripheral nervous system. The

25 promoter/promoters may be derived from promoter sequences of eukaryotic genes. For example, promoters may be derived from the genome of a cell in which expression of the heterologous gene is to occur, preferably a mammalian, preferably human cell of the peripheral nervous system. With respect to eukaryotic promoters, they may be promoters that function in a ubiquitous manner (such as promoters of β -

30 actin, tubulin) or, alternatively, a tissue-specific manner, such as the neuron-specific enolase (NSE) promoter. They may also be promoters that respond to specific stimuli, for example promoters that bind steroid hormone receptors. Viral promoters

may also be used, for example the Moloney murine leukemia virus long terminal repeat (MMLV LTR) promoter or other retroviral promoters, the human or mouse cytomegalovirus (CMV) IE promoter, or promoters of herpes virus gene including those driving expression of the latency associated transcripts.

5 Expression cassettes and other suitable constructs comprising a heterologous gene and control sequences can be made using routine cloning techniques known to persons skilled in the art (see, for example, Sambrook *et al.*, 1989, Molecular Cloning – A Laboratory Manual; Cold Spring Harbor Press).

10 It may also be advantageous for the promoters to be inducible so that the levels of expression of the heterologous gene can be regulated during the life-time of the cell. Inducible means that the levels of expression obtained using the promoter can be regulated. For example, in a preferred embodiment where more than one heterologous gene is inserted into the HSV genome, one promoter would comprise a promoter responsive to the tet repressor/VP16 transcriptional activator fusion protein
15 previously reported (Gossen and Bujard, 1992, Gossen *et al.*, 1995), and driving the heterologous gene the expression of which is to be regulated. The second promoter would comprise a strong promoter (e.g. the CMV IE promoter) driving the expression of the tet repressor/VP16 fusion protein. Thus in this example expression of the first heterologous gene would depend on the presence or absence of
20 tetracycline.

25 In preferred embodiments of the current invention, a promoter is used which allows gene expression during herpes virus latency. Such promoters may be derived from the LAT region (e.g. LAP2; Goins *et al.* 1994) and may be chimaeric promoters containing sequences from the LAT region fused to a LAT or non-LAT promoter or
30 other elements (e.g. Lachmann and Efsthathiou, 1997). These promoters may function from within the endogenous LAT region of the virus or when inserted at a heterologous site in the virus genome. In particularly preferred embodiments chimaeric promoters consisting of a non-LAT promoters such as the CMV, MMLV or NSE promoters, or fragments thereof, linked to LAT-derived sequences, including those present in LAP1 (Goins *et al.* 1994), LAP2 (Goins *et al.* 1994) and/or LAT P2 are used. The LAT P2 region is here defined as HSV1 nucleotides 118866-120219 of HSV strain 17+ (GenBank HE1CG: from PstI-BstXI sites), and includes

fragments or derivatives of this region, including homologous regions of other HSV1 strains and of HSV2 strains, which are capable of providing a long-term expression capability to promoters to which they are linked. Such chimaeric promoters may also allow the expression of multiple heterologous genes by placing
5 pairs of promoters away from a central LAT-derived element such that two genes may be expressed from a single expression cassette (e.g. as in Thomas *et al.* 1999).

Heterologous genes in viruses for therapeutic uses will typically encode polypeptides of therapeutic use. For example, genes which may modulate pain, stimulate nerve re-growth or prevent nerve degeneration. Heterologous genes may
10 be wild-type genes or may be mutated to enhance the function of the gene or to inhibit function of the gene. For example, where an endogenous gene is over expressed in a peripheral nervous system disorder, a non-functional gene which disrupts the function of the endogenous gene may be useful in treating the disorder.

In particular, the heterologous gene/genus may encode a polypeptide capable
15 of modifying responses to painful stimuli or reducing chronic pain, for example a protein capable sequestering, for example nerve growth factor, other pain modulating neurotrophic factor or neurotrophic factor-like molecules, or substance P or other neuropeptides. The heterologous gene/genus may also encode a polypeptide capable of stimulating the regrowth of damaged nerves or preventing the further degeneration
20 of nerves in degenerative conditions.

Suitable heterologous genes include growth factors such as NT-3, NT-4, NT-5, Reg-2, CNTF, GDNF, BDNF, NGF, TrkA and TrkB, neuropeptides such as substance P and galanin, anti-apoptotic genes such as Bcl2 and pain modulators such as enkephalin.

25 Heterologous genes may also include marker genes (for example encoding p-galactosidase or green fluorescent protein or other fluorescent proteins) or genes whose products regulate the expression of other genes (for example, transcriptional regulatory factors including the tet repressor/vmw65 transcriptional activator fusion protein described above).

30 Target validation, gene therapy, and other therapeutic applications may well require the administration of multiple genes. The expression of multiple genes may be advantageous for the treatment of a variety of conditions. Herpes viruses are

uniquely appropriate as they do not have the limited packaging capabilities of other viral vector systems. Thus multiple heterologous genes can be accommodated within its genome. There are, for example, at least two ways in which this could be achieved. For example, more than one heterologous gene and associated control
5 sequences could be introduced into a particular HSV strain either at a single site or at multiple sites in the virus genome. It would also be possible to use pairs of promoters (the same or different promoters) facing in opposite orientations away from each other, these promoters each driving the expression of a heterologous gene (the same or different heterologous gene) as described above.

10 A virus may comprise one, two, three, four or more heterologous genes. Where the gene is a test gene for use in a method of target validation, the virus typically comprises one heterologous gene. The virus may comprise one, two or more copies of a heterologous gene. A virus for use in a method of target validation may comprise more than one heterologous test gene. This will enable several genes,
15 for example from 2 to 20 genes, preferably from 2 or 4 to 10 genes, to be screened simultaneously, or interactions between genes under test to be examined. If an effect of a test gene is observed, further viruses carrying fewer test genes, for example one or two test genes, may be used to determine which of the genes is responsible for the observed effect.

20 Heterologous genes for use in target validation methods will typically be suspected to be of importance in a peripheral nervous system disorder. For example, a gene may be implicated in a disorder due to its presence on region of a chromosome linked to the disorder. Alternatively a gene may be implicated in a peripheral nervous system disorder if it is known to be expressed in the peripheral
25 nervous system. A gene may be implicated in a peripheral nervous system disorder particularly if it is upregulated or down-regulated in a peripheral nervous system cell or in tissue innervated by a peripheral nervous system cell during a natural, artificial or induced disease state or following nerve damage or following administration of a stimuli inducing an acute or chronic pain state or other stimuli which induces a
30 dysfunction in or alteration to the normal functioning of the peripheral nervous system or a peripheral nerve cell.

Monitoring Phenotype

As used herein the term "phenotype" is used to refer to any physical manifestation of a disorder of the central nervous system. The term "phenotype" is thus intended to include changes at the molecular, cellular, tissue, cognitive and
5 behavioural levels. Damage and trauma to the central nervous system, for example as a result of physical and chemical injury is also included within the term "phenotype".

Any suitable phenotype associated with a peripheral nervous system disorder may be monitored. For example, the sensitivity of an animal to pain may be
10 monitored using any suitable assay for monitoring the behavioral response of an animal to pain stimuli. A test gene which influences pain sensitivity will typically enhance or diminish the response of the animal to a given stimulus. Control responses may be determined by testing an animal prior to inoculation with the virus or by testing one or more control animals. A control animal is an animal which has
15 not been inoculated with a virus or which has been inoculated with a control virus which does not express a heterologous gene or which expresses a heterologous gene that does not affect the phenotype of interest. A phenotype associated with a peripheral nervous system disorder may also be monitored at the cellular or molecular level. This is particularly important when monitoring the effect of a test
20 gene *in vitro*, for example following isolation of peripheral nervous system cells which have been transduced *in vivo* following peripheral inoculation with a virus of the invention. For example, the cellular response to the application of a chemical stimulus such as capsaicin may be monitored. This may be achieved using electrophysiological techniques. Alternatively any changes in gene expression
25 within a peripheral nervous system cell in response to the test gene may be monitored, as may responses to stimuli which might otherwise induce cell death.

A test gene may be suitable as a target for gene therapy or for small molecule modulators useful in the treatment of a peripheral nervous system disorder if it influences a phenotype associated with the disorder. A gene identified as a target
30 using a method of the invention may be used in a method of screening for agents useful in the treatment of a peripheral nervous system disorder if it influences a phenotype associated with the disorder. Generally a test gene may be considered to

influence a phenotype if it inhibits or enhances the phenotype, for example expression of a phenotype may be increased or decreased by at least 5%, for example by at least 10%, at least 15%, at least 20% or at least 25%, preferably by at least 30%, for example at least 40% or at least 50%, more preferably by at least 70%, for example, at least 80% or at least 90% compared to controls.

Inoculation of a peripheral nerve

A replication incompetent herpes virus is typically inoculated into a peripheral nerve by direct application to the nerve, for example by injection.

Generally the virus is applied at titres of from 10^4 to 10^{10} pfu, preferably from 10^7 to 10^{10} , more preferably at least 10^8 pfu. Typically from $1\mu\text{l}$ to $20\mu\text{l}$, preferably from $2\mu\text{l}$ to $10\mu\text{l}$, more preferably from $2\mu\text{l}$ to $5\mu\text{l}$ of virus suspension is injected.

Any peripheral nerve, for example the sciatic, femoral, ulnar, median or radial nerves may be injected with the virus. The nerve is preferably a mammalian nerve, for example a rodent such as a mouse or a rat, a primate such as monkey or a weasel or a pig nerve. The nerve may be present in a non-human animal which models a peripheral nervous system disorder. The animal may be a transgenic animal.

Therapeutic uses

Replication incompetent herpes viruses capable of infecting cells of the peripheral nervous system as described herein may be used in methods for treating or preventing peripheral nervous system disorders. As used herein, the term "disorder" is intended to include any condition of the nervous system which affects the normal functioning of the peripheral nervous system. "Conditions", "diseases" and "injury" to the peripheral nervous system are thus encompassed by the term "disorder". The term "disorder" also includes animal models of peripheral nervous system disorders. In particular, viruses of the invention may be used for the treatment of pain states or for the stimulation of nerve regrowth or to prevent nerve degeneration.

The herpes viruses of the present invention may thus be used to deliver therapeutic genes to a human or animal in need of treatment. Delivery of therapeutic genes using the herpes viruses of the invention may be used to treat peripheral

nervous system disorders, for example pain, degenerative conditions or to stimulate nerve regrowth. Generally, the condition of a patient suffering from a peripheral nervous system disorder is improved by administration of the virus.

One method of administering therapy involves inserting a therapeutic gene
5 into the genome of the herpes virus of the invention, as described above, and then combining the resultant recombinant virus, as active ingredient, with a pharmaceutically acceptable carrier or diluent to produce a pharmaceutical composition. Suitable carriers and diluents include isotonic saline solutions, for example phosphate-buffered saline.

10 Gene delivery to cells of the peripheral nervous system may be carried out by direct injection of the vector composition into a peripheral nerve of a human or animal. The amount of virus administered is in the range of from 10^4 to 10^{10} pfu, preferably from 10^5 to 10^8 pfu, more preferably about 10^6 to 10^8 or 10^9 pfu, or from 10^8 to 10^{10} pfu. When injected, typically from 1-200 μ l preferably from 1 to 10 μ l of
15 virus suspension, depending on the species and nerve, in a pharmaceutically acceptable suitable carrier or diluent, is administered.

The routes of administration and dosages described are intended only as a guide since a skilled practitioner will be able to determine readily the optimum route of administration and dosage. The dose of a modulator may be determined according
20 to various parameters, especially according to the substance used; the age, weight and condition of the patient to be treated; the route of administration; and the required regiment. A physician will be able to determine the required dosage for any particular patient.

25 The invention will be described with reference to the following Examples, which are intended to be illustrative only and not limiting.

EXAMPLES

30 Materials and Methods

Virus Strains

Replication incompetent virus strains are derived from HSV1 strain 17+, the

nucleotide sequence of which is deposited in GenBank (Accession No. HE1CG). These can be propagated on 27/12/M:4 cells which are BHK cells containing the HSV ICP27 and ICP4-encoding genes and equine herpes virus gene 12 (Thomas *et al.*, 1999). These cells allow efficient growth of viruses with mutations in ICP4, ICP27 and with mutations in vmw65 reducing its transactivating activity. Viruses not containing mutations in ICP27 but having mutations in ICP4 and vmw65 can be propagated in B4/2 cells which are similar to 27/12/M:4 cells but do not contain the ICP27-encoding gene. Alternatively B130/2 cells can be used, either with or without HMBA, which complement mutations to only ICP27 (Howard *et al.* 1998) for the growth of viruses not including mutations to ICP4.

The current invention was exemplified using virus strain 1764/27-/4-/pR191acZ and 1764/4-pR191acZ. The first virus is deleted for ICP34.5, ICP27 and ICP4 with an inactivating mutation in vmw65 as previously described for virus strain 1764/27-/4-/pR20.5 (Thomas *et al.* 1999). The second virus is identical to the first except that it is not deleted for ICP27. These viruses contains a CMV promoter (from pcDNA3; Invitrogen)/lacZ (pCH110; Pharmacia)/poly A (from pcDNA3) cassette inserted into both copies of the LAT region between the BstXI sites at nts 120,219 and 120,413 rather than the insertion into ICP4 described in Thomas *et al.* 1999. Virus strain 1764/27-/4-/pR19GFP is identical to virus strain 1764/27-/4-/pR191acZ except that the lacZ gene has been directly replaced by the GFP gene (Clontech). Other virus strains 17+/pR191acZ (Wagstaff *et al.* 1998; ICP27 deleted) and 1764/27-/pR20.5/vhs (ICP34.5, ICP27 deleted, inactivating mutation in vmw65) were also used in initial footpad experiments.

Example 1. Replication incompetent HSV gives only low efficiency gene delivery to cells of the peripheral nervous system following footpad inoculation.

Balb/c mice were inoculated by the footpad route as previously described (Coffin *et al.* 1996) with 20µl of virus strains 1764/27-/4-/pR191acZ, 17+/pR191acZ or 1764/27-/pR20.5/vhs at titres of 1×10^7 pfu/ml, 1×10^8 pfu/ml and 5×10^8 pfu/ml. DRGs were removed 1 week after inoculation and fixed and stained with X-gal as previously described (Coffin *et al.* 1996). In contrast to results previously obtained with replication competent viruses with mutations in ICP34.5 or vmw65 and ICP34.5

(Coffin *et al.* 1996), very little gene expression was seen at any of the viruses at any of the titres tested. However with 1764/27-/4-/pR191acZ a maximum of 30 X-gal staining cells was seen at the highest titre used compared to a maximum 5 cells with the less disabled viruses which were also tested. Therefore replication incompetent HSV cannot be used for efficient gene delivery to dorsal root ganglion cells following footpad inoculation in mice although the most highly disabled replication incompetent vector gave slightly improved results compared to the other replication incompetent vectors tested.

Example 2. Genes can be efficiently delivered to cells of the peripheral nervous system using a replication incompetent HSV vector with mutations to ICP27, ICP4 and vmw65 following direct injection into a peripheral nerve.

Balb/c mice were inoculated by direct injection into sciatic nerve with 2-5µl of virus strain 1764/27-/4-/pR191acZ at titres of 1×10^7 pfu/ml, 1×10^8 pfu/ml and 5×10^8 pfu/ml. For inoculation, animals were anaesthetised, the sciatic nerve exposed, and virus suspension injected with a glass micropipette. The wound was then closed and animals allowed to recover. DRGs were removed 1 week after inoculation and fixed and stained with X-gal as above. In contrast to the results above, some gene delivery occurred at the lower two titres (e.g. <20 cells X-gal +ve at 1×10^7 and 1×10^8), and very high gene delivery efficiency could be achieved at a titre of 5×10^8 with many hundreds to thousands of X-gal staining, lacZ+ve, cells being observed. Therefore, at sufficient titre, replication incompetent HSV can be used for highly efficient gene delivery to dorsal root ganglion cells following direct peripheral nerve inoculation of vector suspensions.

25

Example 3. Genes can be efficiently delivered to cells of the peripheral nervous system using a replication incompetent HSV vector with mutations to ICP4 and vmw65 following direct injection into a peripheral nerve.

Balb/c mice were inoculated by direct injection into sciatic nerve with 2-5µl of virus strain 1764/4-/pR191acZ at a titre of 3×10^8 pfu/ml. For inoculation, animals were anaesthetised, the sciatic nerve exposed, and virus suspension injected with a glass micropipette. The wound was then closed and animals allowed to recover.

30

DRGs were removed 1 week after inoculation and fixed and stained with X-gal as above. Similar to the results above in Example 2, many X-gal staining, lacZ +ve cells could be observed.

Example 4. Replication incompetent HSV vectors expressing Cre recombinase
5 **direct efficient excision of intervening gene sequences in loxP containing**
transgenic mice.

Virus strain 1764/27-/4-/pR19Cre was constructed by replacement of the lac
Z gene in virus strain 1764/27-/4-/pR19lacZ with the Cre recombinase gene by
homologous recombination with plasmid pR19Cre. Plasmid pR19Cre was
10 constructed by replacement of the lacZ gene in plasmid pR19 (wagstaff et al 1998)
with the Cre recombinase gene (Sterberg and Hamilton 1981). Modified Rosa26
mice (Mao et al 1999) which are a transgenic line containing lacZ inserted at a
genomic position allowing constitutive expression but preceded by the neomycin
resistance gene flanked by loxP sites which prevents lacZ expression in the absence
15 of prior Cre recombinase mediated recombination were inoculated via the sciatic
nerve with 2-5µl of either virus strain 1764/27-/4-/pR19Cre or virus strain 1764/27-
/4-/pR19GFP at titres of 3×10^8 pfu/ml. 2 days later DRGs were removed and fixed
and stained with X-gal as above after observation for GFP fluorescence by
fluorescence microscopy. DRGs from mice inoculated with virus strain 1764/27-/4-
20 /pR19GFP showed no X gal staining as expected but did show abundant GFP
fluorescence. Thus virus infection per se does not stimulate recombination at the
loxP sites in Rosa26 mice and does not therefore allow lacZ expression. DRGs from
mice inoculated with virus strain 1764/27-/4-/pR19Cre however, while showing no
GFP fluorescence as expected showed abundant lacZ expression, in similar numbers
25 of cells as GFP was expressed following inoculation with virus strain 1764/27-/4-
/pR19GFP, indicating that Cre-mediated recombination at the loxP sites had occurred
and therefore that functional Cre recombinase had been produced.

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CLAIMS

1. Use of a replication incompetent herpes virus comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system disorder by administering said medicament to a peripheral nerve.
2. A use according to claim 1 wherein said virus is a herpes simplex virus 1 or 2.
3. A use according to claim 1 or 2, wherein said virus lacks at least one functional immediate early gene.
4. A use according to any one of claims 1 to 3, wherein said virus lacks at least one functional essential gene which is not an immediate early gene.
5. A use according to claims 3 or 4, wherein said virus lacks a functional VMW65 gene due to a mutation in said gene which abolishes its transcriptional-activation activity.
6. A use according to any one of claims 3 to 5 wherein said immediate early gene is, or genes are, selected from genes encoding ICP0, ICP4, ICP22, ICP27 and ICP47.
7. A use according to claim 5 or 6 wherein said virus lacks both a functional gene encoding ICP4 and a functional gene encoding ICP27 and which has an inactivating mutation in the gene encoding vmw65 abolishing its transcriptional activation activity.
8. A use according to claim 6 wherein said virus lacks functional genes encoding ICP0, ICP4, ICP22 and ICP27.

9. A use according to any one of the preceding claims wherein said virus is a herpes virus amplicon vector.

10. A use according to any one of the preceding claims wherein said
5 heterologous gene is operably linked to a control sequence permitting expression of said heterologous gene in a cell of the peripheral nervous system.

11. A use according to claim 10, wherein said heterologous gene is operably linked to a control sequence active during herpes virus latency.

10

12. A use according to claim 11, wherein said control sequence comprises a fragment of a herpes simplex virus latency associated transcript (LAT) region and non-LAT promoter.

13. A use according to any one of the preceding claims wherein said
15 heterologous gene encodes a polypeptide or antisense RNA of therapeutic use.

14. A use according to claim 13 wherein said polypeptide or RNA is capable of modulating pain, stimulating nerve regeneration or preventing nerve
20 degeneration.

15. A method of treating a subject suffering from a disorder of the peripheral nervous system or of preventing a peripheral nervous system disorder in a subject at risk thereof, which method comprises inoculating a therapeutically
25 effective amount of a replication incompetent herpes virus into a peripheral nerve of the subject.

16. A method according to claim 15 wherein said virus is as defined in any one of claims 2 to 12.

30

17. A method according to claim 15 or 16 wherein said heterologous gene encodes a polypeptide or antisense RNA of therapeutic use.

18. A method according to claim 17 wherein said polypeptide or RNA modulates pain, stimulates nerve regeneration or prevents nerve degeneration.

5 19. An agent comprising a replication incompetent herpes virus for treating a disorder of the peripheral nervous system by administering said agent to a nerve of the peripheral nervous system.

20. A method of determining whether a gene has an effect on a phenotype
10 associated with a peripheral nervous system disorder or on a cell of the peripheral nervous system which is relevant to a peripheral nervous system disorder, which method comprises:

- (i) inoculating a replication incompetent herpes virus comprising a heterologous gene into a peripheral nerve; and
- 15 (ii) monitoring a phenotype of said disorder or an effect of expression of said gene on said cell to determine thereby whether said gene has an effect relevant to said disorder.

21. A method according to claim 20, wherein said cell is a neuron.
20

22. A method according to claim 21 or 22 wherein said gene is expressed in a neuron of the peripheral nervous system.

23. A method according to any one of claims 20 to 22 wherein said virus
25 is as defined in any one of claims 2 to 12.

24. A method according to any one of claims 20 to 23, wherein said gene encodes a protein selected from a neuropeptide and a growth factor.

30 25. A method according to any one of claims 20 to 24, wherein said gene is implicated in a peripheral nervous system disorder.

26. A method according to any one of claims 20 to 25, wherein said disorder is selected from motor neuron disease, chronic pain and peripheral nerve damage.

5 27. A method according to any one of claims 20 to 27, wherein said nerve is *in vivo*.

28. A method according to any one of claims 20 to 28, wherein said nerve is a mammalian nerve.

10

29. A method according to any one of claims 20 to 29, wherein said nerve is from or in a non-human animal which models said disorder.

30. A method according to claim 30 wherein said animal is a
15 transgenic animal.

31. A method according to any of claims 20 to 30 wherein said phenotype is selected from sensitivity to pain stimuli, nerve regeneration and nerve degeneration.

20

32. A method of screening genes implicated in a peripheral nervous system disorder to identify a target for gene therapy or for small molecule modulators, which method comprises a method according to any one of claims 20 to 31.

25

INTERNATIONAL SEARCH REPORT

Inter. Appl. No.
PCT/GB 00/04983

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/869 C12N7/04 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)
EP0-Internal, WPI Data, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WF GOINS ET AL.,: "Herpes simplex virus type 1 vector-mediated expression of nerve growth factor protects dorsal root ganglion neurons from peroxide toxicity" JOURNAL OF VIROLOGY, vol. 73, no. 1, January 1999 (1999-01), pages 519-532, XP002164865 the whole document --- -/--	1-3,6, 10-32

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another claim or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *Z* document member of the same patent family

Date of the actual completion of the international search

23 April 2001

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 00/04983

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>DATABASE BIOSIS [Online] BIOSCIENCES INFORMATION SERVICE, PHILADELPHIA, PA, US; DP WOLFE ET AL.: "Long term systemic delivery of nerve growth factor for treatment of peripheral neuropathies using replication defective genomic HSV vectors" XP002165708 Database accession number= PREV200000139261 abstract & 29th Annual Meeting of the Society for Neuroscience; Miami Beach, FL, USA; October 23-28, 1999</p>	<p>1,2, 13-29, 31,32</p>
X	<p>--- DATABASE BIOSIS [Online] BIOSCIENCES INFORMATION SERVICE, PHILADELPHIA, PA, US; MA LEVATTE ET AL.: "Analysis of a multi-mutant herpes simplex virus type 1 for gene transfer into sympathetic preganglionic neurons and a comparison to adenovirus vectors" XP002164867 Database accession number=PREV199800367232 abstract & Neuroscience, Oct. 1998, Vol. 86 (4), pages 1321-1336</p>	<p>1,2,4, 13-32</p>
X	<p>--- WO 96 27672 A (GLORIOSO JOSEPH C ;FINK DAVID J (US)) 12 September 1996 (1996-09-12)</p>	<p>1-3,6, 10-32</p>
Y	<p>the whole document, in particular page 13 lines 30-34 and page 17 line 35 to page 18 line 8</p>	<p>4,8,9</p>
X	<p>--- WO 98 04726 A (BROWN SUZANNE MOIRA ;MACLEAN ALASDAIR RODERICK (GB); MEDICAL RES C) 5 February 1998 (1998-02-05)</p>	<p>1-3,5-7, 10-32</p>
Y	<p>the whole document, in particular page 10 lines 6-14 and claim 11</p>	<p>4,8,9</p>
X	<p>--- WO 97 20935 A (EFSTATHIOU STACEY ;UNIV CAMBRIDGE TECH (GB); LACHMANN ROBIN HENRY) 12 June 1997 (1997-06-12)</p>	<p>1-3,6, 8-32</p>
	<p>the whole document, in particular page 16 lines 5-7, page 17, page 23-25</p>	
X	<p>--- WO 97 13866 A (UNIV BRITISH COLUMBIA ;CYNADER MAX (CA); TUFARO FRANCIS (CA)) 17 April 1997 (1997-04-17)</p>	<p>1-4,6, 10-32</p>
	<p>the whole document</p>	

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/GB 00/04983

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	J.A. PALMER ET AL.,: "Development and optimization of herpes simplex virus vectors for multiple long-term delivery to the peripheral nervous system" JOURNAL OF VIROLOGY, vol. 74, no. 12, June 2000 (2000-06), pages 5604-5618, XP002164866 the whole document ---	1-3,5-7, 10-32
P,X	DATABASE BIOSIS [Online] BIOSCIENCES INFORMATION SERVICE, PHILADELPHIA, PA, US; J. GOSS ET AL.,: "Protection of spinal motor neurons from degeneration following root avulsion in rats by a HSV vector coding human Bcl-2" XP002165709 Database accession number=PREV200100076066 abstract & 30th Annual Meeting of the Society of Neuroscience; New Orleans, LA, USA; November 04-09, 2000 -----	1-3,6, 10, 13-23, 26-29, 31,32

INTERNATIONAL SEARCH REPORT

International application No.
PCT/GB 00/04983

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 15-18 and 20-32 are directed to methods of treatment and/or diagnostic of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is an immediate early gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

2. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is not an immediate early gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

3. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is an immediate early gene and lacks a functional non-essential gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

4. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is not an immediate early gene and lacks a non-essential gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

virus (vector)

5. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is an immediate early gene and another which is not an immediate early gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

6. Claims: 1-8, 10-32 (partial)

use of a replication incompetent (genomic-based) herpes virus (vector) comprising a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, said use wherein the virus is a herpes simplex virus 1 or 2 and wherein the virus lacks at least one functional essential gene which is an immediate early gene and another which is not an immediate early gene and it further lacks a non-essential gene; method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

7. Claims: 1-32 (partial)

use of a replication incompetent herpes virus (vector) which comprises a heterologous gene in the manufacture of a medicament for use in treating or preventing a peripheral nervous system (PNS) disorder by administering said medicament to a peripheral nerve, wherein the virus is a herpes virus amplicon vector (herpes simplex virus 1 or 2); method of treating a subject suffering from a disorder of the PNS or of preventing a PNS disorder in a subject at risk thereof, which method comprises inoculating a therapeutically effective amount of said replication incompetent (genomic-based) herpes virus (vector) into a peripheral nerve of the subject; an agent comprising said replication incompetent (genomic-based) herpes virus (vector); a method of determining whether a gene has an effect on a phenotype associated with a PNS disorder or on a cell of the PNS which is relevant to a PNS disorder which comprises inoculating said replication incompetent (genomic-based) herpes virus (vector); a method of screening genes implicated in a PNS disorder to identify a target for gene therapy or for small molecule modulators using said replication incompetent (genomic-based) herpes virus (vector)

INTERNATIONAL SEARCH REPORT

Information on patent family members

Inter. Appl. Application No

PCT/GB 00/04983

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